
FINE NEEDLE ASPIRATION—SOLID MASS (Head or neck)

Synonyms

- *Thyroid mass aspiration*
- *Salivary gland aspiration*
- *Parotid mass aspiration*
- *Submandibular gland aspiration*
- *Sublingular gland aspiration*
- *Lymph node aspiration*

Test Code: 9006 / **CPT Code:** 88173

Notes:

1. For multiple nodules on the same patient:
 - Each specimen should have a separate Cytolyt container and requisition card.
 - Clearly label each slide and Cytolyt container with specific specimen source (i.e. right/left, upper/lower).

Supplies:

1. Syringes, optional syringe holder and small gauge needles (not provided by DCCC).
2. Non-sterile specimen container with 30ml of CytoLyt.
3. Yellow Non-Gyn requisition card.
4. Biohazard bag.
5. Plastic transport container with glass slides with frosted ends.
6. Coplin jar with 95% methanol or cytology spray fixative.

Collection:

1. USE UNIVERSAL PRECAUTIONS AND BE CAREFUL TO AVOID NEEDLE STICKS.
2. Label all slides with patient's name in pencil on the frosted end of the slides.
3. Write "Air dry" on the top of two of the slides and "Fix" on the other two.
4. Attach needle to syringe.
5. Isolate mass with fingers.
6. Insert needle into mass.
7. Pull back on syringe plunger to create negative pressure.
8. Gently move needle back and forth within the mass in an up and down motion while changing angles. Do not break skin barrier.
9. Material should go no farther than the needle hub. Once material is visualized in the hub (either blood or cellular material) the aspiration is stopped.
10. Release the syringe plunger to equalize the pressure and withdraw the needle. Do not push the plunger.
11. Prepare slides:
 - Place two slides on the work surface. (One should be marked "Air dry" and the other as "Fix".)
 - Place 2-3 drops of FNA material near the center of one slide.
 - Cover droplets with the face of a second plain glass slide.
 - As the droplets spread between the face to face slides, very gently pull the slides across each other in opposite directions.
 - The "Fix" slide should be IMMEDIATELY fixed for 30 seconds in a Coplin jar of 95% alcohol or spray slides with cytology fixative.
 - Place the "Air dry" slide face up on a paper towel to dry.
 - After fixation, "Fix" slides should also be allowed to dry, face up, on a paper towel.
 - Place slides in transport container.
12. The remainder of the FNA specimen should be rinsed into a container of CytoLyt.
13. Rinse needle and syringe in CytoLyt by placing needle in pre-filled specimen container.

14. Pull back on syringe until CytoLyt is drawn into the syringe and express it back out into the container.
 15. Repeat steps 4-14 for second pass.
 16. Use a clean needle for each pass.
 17. Multiple passes from the same site should be rinsed in one specimen container.
 18. Passes from different sites must be rinsed in separate specimen containers and require separate requisition cards.
 19. Any subsequent passes should be rinsed directly into the CytoLyt container.
 20. Secure the lid and label specimen container with the following:
 - Patient's name
 - Physician's name
 - Source of specimen
 - Date of collection
 21. Complete the yellow Non-Gyn requisition card with the following information:
 - Patient's name, birth date and sex
 - Physician's name and clinic account number
 - Source of specimen
 - Date of collection
 - All pertinent clinical information (especially history of cancer, chemotherapy and radiation treatments).
 - A separate requisition card must be filled out for each specimen.
 22. Send specimen container, slides and requisition card in a Biohazard bag to DCCC.
 - Tighten lid of CytoLyt container.
 - Place CytoLyt and slide container in the ziplock portion of the bag and seal it.
 - Place the yellow Non-Gyn requisition card into the back pocket with the overlapping flaps.
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Storage Instructions:

1. If more than a 24-hr delay is anticipated between collection and receipt in the laboratory, please refrigerate specimen.
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